STRUCTURE OF THE NEUTRAL CARBOHYDRATE SIDE-CHAINS IN ANTI-COMPLEMENTARY ACIDIC POLYSACCHARIDES FROM THE ROOT OF Angelica acutiloba KITAGAWA*

HIROAKI KIYOHARA AND HARUKI YAMADA[†]

Oriental Medicine Research Center of the Kitasato Institute, Minato-ku, Tokyo 108 (Japan)
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ABSTRACT

The endo- α -(1 \rightarrow 4)-polygalacturonase-resistant regions (PG-1a–1c) from three anti-complementary acidic polysaccharides (AR-2IIa-IIc), isolated from the root of Angelica acutiloba Kitagawa, were subjected to base-catalysed β -elimination in the presence of sodium borodeuteride. Each PG-1 gave a neutral (N) and an acidic fraction (A), but each fraction A was resistant to further β -elimination. Each fraction N consisted of Ara, Gal, and Rha, whereas each fraction A contained a large proportion of GalA and small proportions of neutral sugars. Methylation and g.l.c.-m.s. analysis showed that each fraction N contained Gal- $(1\rightarrow 4)$ -Rha-ol-1-d, Gal- $(1\rightarrow 4)$ -Gal-ol-1-d, Gal- $(1\rightarrow 6)$ -Gal-ol-1-d, Gal- $(1\rightarrow 6)$ -Gal- \rightarrow Rha-ol-1-d, and Gal- $(1\rightarrow 6)$ -Gal- $(1\rightarrow 6)$ -Gal-ol-1-d. Methylation analysis suggested that fraction Na from AR-2IIa contained a high-molecular-weight galactan with Gal and Ara side-chains, and that fraction Aa from AR-2IIa consisted mainly of 4-linked GalA together with small amounts of Gal. Degradation of fraction Aa with lithium in ethylenediamine gave $Gal-(1\rightarrow 6)$ -Gal-ol-1d, $Gal\rightarrow Gal$ -ol-1d, Gal-(1 $\rightarrow 6$)-Gal-ol, and Gal \rightarrow Gal-ol. AR-2IIa and PG-1a reacted with β -glucosyl-Yariv antigen, whereas AR-2IIb-IIc and PG-1b-1c reacted with the antigen weakly or negligibly.

INTRODUCTION

Four pectin-like polysaccharides (AR-2IIa–IId), isolated¹ from the root of Angelica acutiloba Kitagawa (Japanese name, Yamato-Tohki), comprise large proportions of polygalacturonan regions and small proportions of "ramified" regions containing the rhamnogalacturonan core and neutral carbohydrate chains that contain¹ mainly Ara and Gal.

Anti-complementary pectic polysaccharides have been isolated from some medicinal plants²⁻⁶ and it is suggested that the neutral carbohydrate side-chains,

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[†]Author for correspondence.

such as arabino-3,6-galactan, contribute to the expression of the activity^{3,6} The "ramified" regions from AR-2IIa–IId express similar activities that are higher than those of the original polysaccharides¹, and we now report on the structure of the neutral carbohydrate side-chains.

EXPERIMENTAL

Materials. — The root of A. acutiloba Kitagawa was purchased from Tochimoto Tenkaidoh (Japan), and the anti-complementary acidic polysaccharides (AR-2IIa–IId)¹ were isolated by hot-water extraction, precipitations with ethanol and Cetavlon (cetyltrimethylammonium bromide), and anion-exchange chromatography. The α- and β-glucosyl-Yariv antigens were gifts from Dr. A. E. Clarke (Plant Cell Biology Research Center, School of Botany, University of Melbourne). Sephadex G-10, G-50, and LH-20, and DEAE-Sephadex A-25 were purchased from Pharmacia, and Bio-gel P-2 (200–400 mesh) and P-4 (–400 mesh) from Bio-Rad. Pectinase from Aspergillus niger was purchased from Sigma, and endo-α-(1→4)-polygalacturonase was purified⁷ by Q.-P. Gao in our laboratory.

General. — Total carbohydrate and uronic acid in column eluates were assayed by the phenol–sulphuric acid⁸ and m-hydroxybiphenyl methods⁹, respectively. Samples were hydrolysed with 2M trifluoroacetic acid for 1.5 h at 121°, and t.l.c. of the hydrolysates was performed on cellulose, using ethyl acetate–pyridine–acetic acid–water (5:5:1:3) and detection with alkaline silver nitrate. Neutral sugars and uronic acids were converted¹⁰ into alditol acetates and analysed³ by g.l.c.

Preparation of "ramified" regions (PG-1a-1c) from AR-2IIa-IIc. — After desterification of AR-2IIa-IIc severally in 0.2M sodium hydroxide for 2 h at room temperature, the products were digested with endo- α -(1 \rightarrow 4)-polygalacturonase⁷ in 50mM acetate buffer (pH 4.2) at 37° for 4 days, then fractionated on Sephadex G-50. The "ramified" regions (PG-1a-1c) were eluted in the void volume.

Gel-diffusion experiments. — AR-2IIa–IId and PG-1a–1c were tested for reactivity to the Yariv antigen by single radial gel-diffusion¹¹. β -Glucosyl-Yariv antigen [1,3,5-tri-(4- β -D-glucopyranosyloxyphenylazo)-2,4,6-trihydroxybenzene] was used as the positive Yariv antigen and α -glucosyl-Yariv antigen as the negative antigen. Samples (15 μ g) were applied to agarose plates containing Yariv antigen (10 μ g/mL) and incubated overnight at room temperature.

Depolymerisation of PG-1a-1c by base-catalysed β -elimination in the presence of sodium borodeuteride. — Each PG-1 (10 mg) was methyl-esterified with diazomethane, then subjected to a base-catalysed β -elimination reaction (0.1M NaOH, 3 h, 100°) in the presence of sodium borodeuteride¹². The procedure was repeated five times and the final products were fractionated¹² on DEAE-Sephadex A-25 (HCOO⁻ form) to give neutral (Na-Nc) and acidic fractions (Aa-Ac).

Treatment¹³ of fraction Aa with lithium in ethylenediamine. — To a solution of fraction Aa in ethylenediamine was added lithium, the mixture was stirred for 1 h at room temperature and then cooled in ice, and water was added to stop the

reaction. The solvent was evaporated, and the products were reduced with sodium borohydride and desalted with AG50W-X8 (H⁺) resin.

Methylation analysis. — Each sample was methylated (Hakomori¹⁴) and the products were purified¹⁵ using a Sep-pak C₁₈ cartridge (Waters Assoc.). The methylated fraction Na was eluted from Sephadex LH-20 (chloroform-methanol, 1:1) to give fractions of high (HMW) and low molecular weight (LMW). Uronic acid in the methylated sample was reduced¹⁵ with sodium borodeuteride in tetrahydrofuran-ethanol (7:3). The methylated product was hydrolysed with 2M trifluoroacetic acid for 1.5 h at 121°, and the products were converted into the corresponding alditol acetates and analysed³ by g.l.c. and g.l.c.-m.s. on an SPB-1 capillary column (SUPELCO).

G.l.c.-m.s. of methylated oligosaccharide-alditols. — The methylated oligosaccharide-alditols obtained from fraction Na-Nc were analysed¹⁶ by g.l.c.-m.s. on an SPB-1 capillary column. C.i.-¹⁷ and e.i.-m.s. fragment ions [A, J, and alditol (ald)]¹⁸ were used to determined the structures.

RESULTS

Release of the neutral carbohydrate side-chains from the "ramified" regions of the acidic polysaccharides by base-catalysed β -elimination in the presence of sodium borodeuteride. — The "ramified" regions (PG-1a-1d) from the anti-complementary polysaccharides (AR-2IIa-IId) were each suggested to consist¹ of a rhamnogalacturonan core to which neutral carbohydrate side-chains were attached. The side chains attached to position 4 of GalA residues can be released¹² from the acidic core as alditol-1-d derivatives by a base-catalysed β -elimination reaction in the presence of sodium borodeuteride without methyl-etherification of polysaccharides.

Because insufficient PG-1d was available, only PG-1a-1c were studied. The products of each β -elimination reaction were eluted from DEAE-Sephadex A-25 (HCOO⁻ form), to give a minor neutral fraction (Na-Nc) and a major acidic fraction (Aa-Ac) (data not shown). Each fraction N comprised Rha, Ara, and Gal (molar ratios of 0.2:0.7:1.0 for Na, 0.2:6.1:1.0 for Nb, and trace:5.2:1.0 for Nc), whereas each fraction A contained small amounts of Rha, Ara, and Gal (molar ratios of trace:0.1:1.0 for Aa, trace:0.8:1.0 for Ab, and trace:0.3:1.0 for Ac) in addition to a large amount of GalA. When fractions Aa-Ac were each subjected to β -elimination, most of the material was recovered as an acidic fraction (data not shown).

Analysis of fraction Na. — Fraction Na was methylated and the products were separated into fractions of high (HMW) and low molecular weight (LMW) by chromatography on Sephadex LH-20. Methylation analysis (Table I) of HMW gave mainly 1,4-di-O-acetyl-2,3,5-tri-O-methyl-, 1,4,5-tri-O-acetyl-2,3-di-O-methyl-, and 1,3,4,5-tetra-O-acetyl-2-O-methyl-arabinitol, and 1,5-di-O-acetyl-2,3,4,6-tetra-O-methyl-, 1,5,6-tri-O-acetyl-2,3,4-tri-O-methyl-, and 1,3,5,6-tetra-O-acetyl-2-acetyl-2,3,4-tri-O-methyl-, and 1,3,5,6-tetra-O-acetyl-2-acetyl

TABLE I

METHYLATION ANALYSIS OF PRODUCTS (PG-1a, Na, HMW, LMW, and Aa) DERIVED FROM AR-2IIa

Glycosyl	Deduced	Composition (mol. %)								
residue	glycosidic linkage	PG-1a	Na	HMW	LMW	Aa				
Ara	terminal (furanosyl)	4.0	9.4	4.6	12.1	5.2				
	terminal (pyranosyl)	1.7	2.8	0.3	2.0	n.d.				
	3- (pyranosyl)	3.2	2.5	1.2	6.8	n.d.				
	4- or 5-	3.4	6.8	6.1	10.4	n.d.				
	3,4- or 3,5-	1.0	6.4	4.4	4.2	n.d.				
Gal	6- (reducing terminal)	$\mathrm{n.d.}^a$	trace	n.d.	trace	trace				
	terminal	4.4	18.9	10.5	22.1	10.8				
	4-	3.2	6.0	1.7	7.4	n.d.				
	3-	3.4	7.0	5.7	8.5	5.9				
	6-	10.7	23.6	47.5	12.5	56.0				
	2,4-	6.0	trace	trace	n.d.	2.7				
	4,6-	6.5	6.8	2.0	1.5	n.d.				
	3,6-	21.3	7.0	11.6	2.0	8.0				
	3,4,6-	20.8	trace	1.4	0.4	n.d.				
Rha	terminal	1.8	trace	0.6	2.2	n.d.				
	2-	1.7	n.d.	trace	trace	n.d.				
	2,4-	4.2	n.d.	n.d.	n.d.	n.d.				
Glc	terminal	0.3	trace	1.7	2.2	n.d.				
Xyl	3,4-	1.8	trace	0.8	1.8	n.d.				
GalA	terminal					n.d.				
	4-					11.3				

aNot detected.

2,4-di-O-methyl-galactitol, whereas LMW gave mainly 1,4-di-O-acetyl-2,3,5-tri-O-methyl- and 1,4,5-tri-O-acetyl-2,3-di-O-methyl-arabinitol, 1,5-di-O-acetyl-2,3,4,6-tetra-O-methyl- and 1,5,6-tri-O-acetyl-2,3,4-tri-O-methyl-galactitol, and a trace of 6-O-acetyl-1,2,3,4,5-penta-O-methylgalactitol-1-d.

Methylated oligosaccharide-alditols of LMW were analysed by g.l.c.-m.s. (Tables II and III). The methylated disaccharide-alditols 1–3 (Table II) gave fragment ions due to non-reducing termini (bA_1 and bA_2) and alditols (aJ_1 and aJ_2), indicating that 1 had a hexosyl—6-deoxyhexitol-1-d unit, and 2 and 3 had hexosyl—hexitol-1-d units. The products 1–3 were eluted from an SPB-1 capillary column in this order and they also gave fragment ions due to the ald series. From the results of e.i.-m.s. and the comparison of retention time with those of standard disaccharide-alditols, 1–3 were identified as $Gal-(1\rightarrow 4)$ -Rha-ol-1-d, $Gal-(1\rightarrow 4)$ -Gal-ol-1-d, and $Gal-(1\rightarrow 6)$ -Gal-ol-1-d, respectively. Of the methylated trisaccharide-alditols (4 and 5) in LMW (Table III), 4 gave fragment ions at m/z 219 and 187 due to the non-reducing terminus (bA_1 and bA_2 , respectively), 266 and 206 due to alditol (aJ_1 and aJ_2 , respectively), and 391 (bcA_2 produced by elimination of the alditol from the trisaccharide-alditol). Therefore, 4 was considered to be $Gal\rightarrow Gal\rightarrow Rha$ -ol-1d. However, the fragment ions assigned to the ald series were not ob-

served, and the glycosidic linkages of the alditol portion could not be deduced. Likewise, the derivative 5 was considered to be $Gal \rightarrow Gal \rightarrow Gal - ol-1-d$. Methylation analysis showed that LMW contained a large amount of 6-linked Gal in addition to 6-linked galactitol-1-d; therefore, 4 and 5 were assumed to be $Gal - (1 \rightarrow 6) - Gal \rightarrow Rha - ol-1-d$ and $Gal - (1 \rightarrow 6) - Gal - (1 \rightarrow 6) - Gal - ol-1-d$, respectively. Thus, HMW consists mainly of $(1 \rightarrow 6)$ -linked galactosyl chains possessing some Gal and Ara side-chains, whereas LMW contained galacto-oligosaccharides with either Rha or Gal at the reducing termini. It was also concluded that PG-1a contained short arabinosyl side-chains consisting of terminal Araf and 4- or 5-linked Ara.

Analysis of fraction Aa. — Fraction Aa was eluted as a broad peak from Bio-gel P-4 in 50mM acetate buffer (pH 5.6) (Fig. 1A). Methylation analysis (Table I) showed that fraction Aa contained mainly Gal and a low proportion of GalA. Since incomplete reduction of GalA in methylated polysaccharides by lithium aluminium hydride has been observed¹⁹, the loss of GalA in the present study is attributed to incomplete reduction of GalA by sodium borodeuteride. It was shown that fraction Aa contained terminal and 6-linked Gal and 4-linked GalA. Only 6-linked galactitol-1-d was formed from Aa.

TABLE II

DIAGNOSTIC IONS ON E.I.-M.S. OF METHYLATED DISACCHARIDE-ALDITOLS IN FRACTION $N_{\rm a}$

Product	Fragm	ent ions	Structure						
	aJ_1	a J 2	bA_I	bA_2	ald				
1	266 (4.1)	206 (51.0)	219 (17.2)	187 (91.7)	319 (0.3)	275 (2.7)	134 (30.2)		Gal- $(1\rightarrow 4)$ -Rha-ol- $1-d$
2	296 (2.5)	236 (27.2)	219 (17.4)	187 (83.4)	362 (5.3)	349 (1.1)	305 (0.4)	134 (21.0)	Gal- $(1\rightarrow 4)$ -Gal-ol- $1-d$
3		236 (20.7)	219 (56.0)	187 (55.3)	337 (0.6)	305 (0.3)	178 (3.8)	146 (20.6)	Gal- $(1\rightarrow 6)$ -Gal-ol- I - d

TABLE III

DIAGNOSTIC IONS ON E.I.-M.S. OF METHYLATED TRISACCHARIDE-ALDITOLS IN FRACTION Na

Product	Fragmer	nt ions [m/z (Structure			
	aJ_1	aJ_2	bA_1	bA_2	bcA_2	
4	266 (2.7)	206 (30.3)	219 (15.5)	187 (82.4)	391 (1.2)	Gal→Gal→Rha-ol-1-d
5	296 (2.6)	236 (6.9)	219 (20.7)	187 (97.9)	391 (1.3)	$Gal \rightarrow Gal \rightarrow Gal - ol - 1 - d$

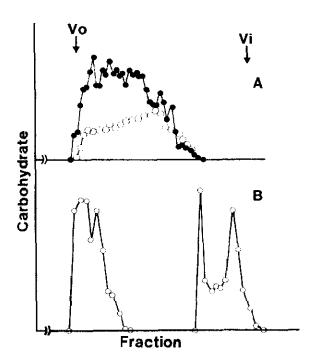


Fig. 1. Gel filtration on Bio-gel P-4 of A, fraction Aa in 50mm acetate buffer (pH 5.6); B, lithium-treated fraction Aa: ○, carbohydrate (490 nm); ●, uronic acid (520 nm); Vo, void volume; Vi, included volume.

Treatment of fraction Aa with lithium in ethylenediamine¹³ decomposed the GalA. Elution of the products from Bio-gel P-4 gave Li-A-1 in the void volume, and lower oligosaccharides Li-A-2 (Fig. 1B). Li-A-1 consisted mainly of Gal, but further analysis could not be performed because of the small amount available. Li-A-2 was methylated (Hakomori¹⁴), and the products were analysed by g.l.c.m.s. C.i.-m.s. (Table IV) gave two protonated molecular ions at m/z 442 and 441, and fragment ions due to aJ_1 (m/z 266 and 265), aJ_2 (m/z 206 and 205), aJ_2OH_2 (m/z) 224 and 223), and bA₁ and bA₂ (m/z) 219 and 187). These results suggested the presence of a hexosyl \rightarrow 6-deoxyhexitol-1-d (6) and a hexosyl \rightarrow 6-deoxyhexitol (7). In e.i.-m.s. (Table V), 6 gave fragment ions of the ald series at m/z 395, 319, 307, and 134, and 6 was assigned as Gal- $(1\rightarrow 4)$ -Rha-ol-1-d, whereas 7 gave fragment ions at m/z 395, 305, and 293, and was assigned as Gal-(1 \rightarrow 2)-Rha-ol. E.i.-m.s. also revealed products 8-10, which were not detected in c.i.-m.s. because of their small amounts. Product 8 gave fragment ions due to aJ_2 at m/z 236 and 235, and to bA_1 and bA_2 at m/z 219 and 187, suggesting the presence of the disaccharidealditols 8a and 8b. In e.i.-m.s., 8a gave fragment ions due to ald series at m/z 337, 178, 146, and 134, whereas **8b** gave fragment ions at m/z 337, 177, 145, and 133, indicating the former to be Gal- $(1\rightarrow 6)$ -Gal-ol-I-d and the latter to be Gal- $(1\rightarrow 6)$ -Gal-ol. Product 9 was eluted in the region for a hexosyltrisaccharide-additol in g.l.c. on an SPB-1 capillary column, and gave fragment ions due to abJ_2 at m/z 439, and to aJ_2 and bA_1 at m/z 235 and 219, suggesting the structure $Gal \rightarrow Gal \rightarrow Gal$ -ol. However, the linkage of the alditol portion could not be deduced because the fragment ions due to ald were not observed. Product 10 was also eluted in the region for a hexosyltrisaccharide-alditol and gave the fragment ions due to abJ_2 (m/z 440), bA_1 (m/z 219), and aJ_2 (m/z 236), indicating the structure $Gal \rightarrow Gal \rightarrow Gal \rightarrow I-d$.

TABLE IV DIAGNOSTIC IONS ON C.I.-M.S. OF METHYLATED OLIGOSACCHARIDE-ALDITOLS IN Li-A-2 DERIVED FROM FRACTION Aa by Lithium-mediated degradation

Product	Fragment	Fragment ions [m/z (relative abundance)]									
	(M+H)+	(M+H)+ -MeOH	aJ_{I}	aJ_2	aJ ₂ OH ₂	<i>bA</i> ₁	bA_2	- alditol			
6	442	410	266	206	224	219	187	hexosyl→			
	(15.9)	(74.6)	(7.3)	(12.1)	(56.6)	(81.3)	(100)	6-deoxyhexitol- <i>1-d</i>			
7	441	409	265	205	223	219	187	hexosyl→			
	(6.9)	(29.4)	(3.5)	(6.7)	(23.9)	(81.3)	(100)	6-deoxyhexitol			

TABLE V

DIAGNOSTIC IONS ON E.I.-M.S. OF METHYLATED OLIGOSACCHARIDE-ALDITOLS IN Li-A-2 DERIVED FROM FRACTION Aa BY LITHIUM-MEDIATED DEGRADATION

Product	Fragme	ent ions	[m/z (rel	ative abi	undance)]			Structure	
	aJ_1	aJ_2	bA_I	bA_2	ald					
6	266 (8.3)	206 (5.4)	219 (29.8)	187 (100)	395 (0.2)	319 (0.3)	307 (0.7)	134 (25.6)	Gal-(1→4)-Rha-ol-1-d	
7	265 (4.8)	205 (3.3)	219 (29.8)	187 (100)	395 (0.2)	305 (1.2)	293 (0.7)		Gal-(1→2)-Rha-ol	
8a		236 (26.0)	219 (23.2)	187 (91.3)	337 (4.0)	178 (3.1)	146 (18.2)	134 (6.7)	Gal- $(1\rightarrow 6)$ -Gal-ol- 1 - d	
8b		235 (56.7)	219 (23.2)	187 (91.3)	337 (4.0)	177 (11.9)	145 (67.8)	133 (60.4)	Gal-(1→6)-Gal-ol	
	aJ_2	abJ_2	bA_I	bA_2	bcA_1	bcA_2				
9	235 (56.9)	439 (12.5)	219 (21.1)	187 (100)					Gal→Gal-ol	
10	236 (36.2)	440 (2.0)	219 (10.4)	187 (100)					Gal→Gal-ol-1-d	

Analysis of fractions Nb, Nc, Ab, and Ac. — Fractions Nb and Nc were also analysed as described above. However, the methylated products were not fractionated on Sephadex LH-20 because the amounts available were too small.

When each methylated fraction N was subjected to g.l.c.-m.s., the same diand tri-saccharide-alditols were detected as in fraction Na (Tables II and III). Methylation analysis (Table VI) showed that Nb and Nc contained mainly terminal Araf, 4- or 5-linked and 3,4- or 3,5-disubstituted Ara, and fraction Nb also contained

TABLE VI					
METHYLATION ANALYSIS OF PG-1b,	PG-1c, AND FRACTIONS	<i>N</i> b, <i>N</i> c,	Ab, A	AND.	Аc

Glycosyl residue	Deduced	Composition (mol. %)								
	glycosidic linkage	AR-2IIb			AR-2IIc					
		PG-1b	Nb	$\mathbf{A}b$	PG-1c	Nc	Ac			
Ara	terminal (furanosyl)	6.3	11.1	3.8	8.5	25.5	3,5			
	terminal (pyranosyl)	2.4	0.7	n.d.	2.7	n.d.	n.d.			
	3- (pyranosyl)	9.4	n.d.	n.d.	13.4	n.d.	n.d.			
	4- or 5-	9.9	26.1	3.3	12.4	28.9	n.d.			
	3,4- or 3,5-	3.3	26.8	n.d.	6.1	25.5	n.d.			
Gal	6- (reducing terminal)	$n.d.^a$	trace	n.d.	n.d.	trace	n.d.			
	terminal	8.8	6.7	9.0	10.6	4.0	3.7			
	4-	3.9	12.6	3.7	4.8	8.3	3.9			
	3-	3.9	2.6	n.d.	3.4	1.3	10.7			
	6-	6.3	8.9	40.5	3.5	5.2	9.5			
	2,4-	10.7	n.d.	n.d.	7.3	trace	4.1			
	4,6-	4.3	n.d.	n.d.	3.2	trace	n.d.			
	3,6-	8.9	4.6	5.6	4.3	1.3	6.4			
	3,4,6-	2.8	n.d.	n.d.	1.2	trace	n.d.			
Rha	terminal	4.5	n.d.	n.d.	4.4	n.đ.	n.d.			
	2-	4.1	n.d.	n.d.	3.4	n.d.	n.d.			
	2,4-	6.6	n.d.	n.d.	7.8	n.d.	n.d.			
Glc^b	terminal	1.0	n.d.	n.d.	1.2	trace	n.d.			
Xyl^b	3,4-	3.0	n.d.	n.d.	2.0	trace	n.d.			
GalA	terminal			n.d.			15.0			
	4-			34.0			43.2			

^aNot detected. ^bNot detected in the original polysaccharides.

mainly 4-linked Gal. A trace of 6-O-acetyl-1,2,3,4,5-penta-O-methylgalactitol-1-d was detected. However, some glycosyl residues, such as terminal and 3-linked Arap and various types of Gal, were decreased or lost in each fraction N by comparison with each PG-1 of AR-2IIb and IIc, possibly by a "peeling" reaction during the β -elimination reaction.

Methylation analysis of the fractions Ab and Ac indicated (Table VI) that fraction Ab contained a large proportion of 4-linked GalA together with terminal and 6-linked Gal, and that fraction Ac contained a large proportion of terminal and 4-linked GalA together with 3- and 6-linked Gal.

Analysis of the $(1\rightarrow 3,6)$ -galactan moiety by using the Yariv antigen. — The β -glucosyl-Yariv antigen reacts²⁰ with $(1\rightarrow 3,6)$ - β -galactan to form a red dye, and $(1\rightarrow 3,6)$ - β -galactan can be quantified¹¹ by single radial gel-diffusion with the β -glucosyl-Yariv antigen. When AR-2IIa-IId were tested by single radial gel-diffusion with the β -glucosyl-Yariv antigen (Fig. 2), AR-2IIa reacted slightly, whereas AR-2IIb-IId did not react. PG-1a reacted strongly with the antigen, whereas PG-1b reacted slightly, and PG-1c did not react. Each polysaccharide and PG-1 did not

react with α -glucosyl-Yariv antigen as the negative control²⁰ (data not shown). These results suggested that AR-2IIa contained a large proportion of a $(1\rightarrow3,6)$ - β -galactan moiety, whereas AR-2IIb-IId contained small or negligible proportions.

DISCUSSION

Acidic pectic polysaccharides^{21,22} are considered to contain a polygalacturonan region and a rhamnogalacturonan as the acidic core with neutral carbohydrate chains attached to position 4 of 2,4-disubstituted Rha in the rhamnogalacturonan core to form the "ramified" region. However, the details of structure of the neutral carbohydrate side-chains and their mode of attachment have not yet been clarified.

The present results suggested that the short galactosyl oligosaccharide chains in the neutral carbohydrate side-chains released from the "ramified" regions of AR-2IIa by the β -elimination reaction are attached to position 4 of GalA residues through 4-linked Rha, and 4- and 6-linked Gal as shown in Table VII. Some pectic polysaccharides contain^{4,16} the sequence \rightarrow 4)-GalA-(1 \rightarrow 4)-Rha-(1 \rightarrow , and it is suggested that short galactosyl chains in AR-2IIa–IIc might be linked to such a chain either as $(Gal)_n\rightarrow Gal-(1\rightarrow 4)$ -GalA-(1 \rightarrow 4)- $[\rightarrow 2)$ -Rha-(1 \rightarrow] or $(Gal)_n\rightarrow Gal-(1\rightarrow 4)$ - $[\rightarrow 2)$ -Rha-(1 \rightarrow]. AR-2IIa also contained long galactosyl chains possessing some Gal and Ara side-chains which were assumed to be attached to rhamnogalacturonan as described above. The long galactosyl chains in AR-2IIa only were also assumed to possess a (1 \rightarrow 3,6)-galactan moiety because AR-2IIa reacted with the β -glucosyl-Yariv antigen. The result of methylation analysis of LMW suggested that AR-2IIa also contained arabino-oligosaccharide chains. It was also assumed that the "ramified" regions of AR-2IIb and IIc might contain more arabinose-rich side-chains than that of AR-2IIa.

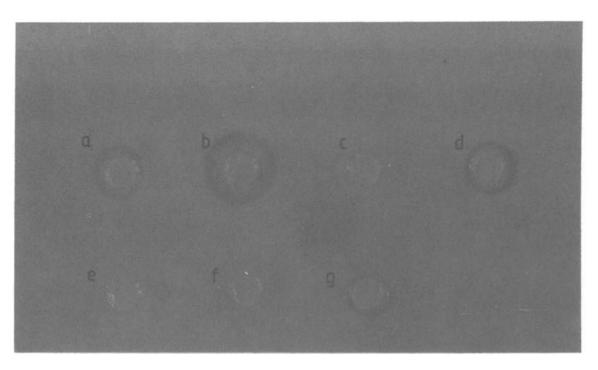


Fig. 2. Single radial gel-diffusion with Yariv antigen: (a) AR-2IIa, (b) PG-1a, (c) AR-2IIb, (d) PG-1b, (e) AR-2IIc, (f) PG-1c, (g) AR-2IId.

TABLE VII

PARTIAL STRUCTURES IN AR-2IIa-IIc

Recently, Lau et al. reported²³ that Rhamnogalacturonan I (RG-I) contained various galacto- and arabino-oligosaccharide side-chains similar to those in AR-2IIa.

The base-catalysed β -elimination reaction of the "ramified" regions from AR-2IIa-IIc gave a large proportion of an acidic fraction (Aa-Ac), that contained galacto-oligosaccharides. Because some of the galacto-oligosaccharides possessed galactitol at the reducing termini, fraction A was produced by incomplete β -elimination of GalA. However, since some galacto-oligosaccharides possessing galactitol-1-d were also detected in the products from fraction Aa after the degradative reaction using lithium, it is suggested that they were attached originally to GalA as in $\rightarrow 4$)-GalA- $(1\rightarrow (Gal)_n$ - $(1\rightarrow 4)$ -GalA- $(1\rightarrow ...)$

Although the structures of the neutral side-chains in the "ramified" region of AR-2IIa were different from those of AR-2IIb and IIc, each "ramified" region showed¹ similar anti-complementary activity. A pectin²⁴ from Zyziphus jujuba was shown to have²⁵ no anti-complementary activity, and to contain²⁴ mainly 4-linked Gal as the neutral sugar. Therefore, the neutral carbohydrate chains possessing the common structure of AR-2IIa–IIc might play a role in the expression of anti-complementary activities of their "ramified" regions. The relationship between the neutral carbohydrate side-chains and anti-complementary activity of "ramified" regions in these polysaccharides is being studied further.

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REFERENCES

- 1 H. KIYOHARA, J.-C. CYONG, AND H. YAMADA, Carbohydr. Res., 182 (1988) 259-275.
- 2 H. YAMADA, Y. OTSUKA, AND S. OMURA, Planta Medica, (1986) 311-314.
- 3 H. YAMADA, H. KIYOHARA, J.-C. CYONG, AND Y. OTSUKA, Carbohydr. Res., 159 (1987) 275-291.
- 4 H. YAMADA, H. KIYOHARA, AND Y. OTSUKA, Carbohydr. Res., 170 (1987) 181-191.
- 5 H. YAMADA. S. YANAHIRA, H. KIYOHARA, J.-C. CYONG, AND Y. OTSUKA, *Phytochemistry*, 26 (1987) 3269–3275.

- 6 Q.-P. GAO, H. KIYOHARA, J.-C. CYONG, AND H. YAMADA, Carbohydr. Res., 181 (1988) 175-187.
- 7 J. F. Thibault and C. Mercier, J. Solid-Phase Biochem., 2 (1977) 295-304.
- 8 M. Dubois, K. A. Gilles, J. K. Hamilton, P. A. Rebers, and F. Smith, *Anal. Chem.*, 28 (1956) 350-356.
- 9 N. BLUMENKRANTZ AND G. ASBOE-HANSEN, Anal. Biochem., 54 (1973) 484-489.
- 10 T. M. JONES AND P. ALBERSHEIM, Plant Physiol., 49 (1972) 926-936.
- 11 G.-J. HOLST AND A. E. CLARKE, Anal. Biochem., 148 (1985) 446-450.
- 12 H. KIYOHARA AND H. YAMADA, Carbohydr. Res., 187 (1989) 117-129.
- 13 J. M. LAU, M. McNeil, A. G. DARVILL, AND P. ALBERSHEIM, Carbohydr. Res., 168 (1987) 219-243.
- 14 S. HAKOMORI, J. Biochem. (Tokyo), 55 (1964) 205-208.
- 15 T. Z. WAEGHE, A. G. DARVILL, M. McNeil, and P. Albersheim, *Carbohydr. Res.*, 123 (1983) 281–304.
- 16 H. KIYOHARA, H. YAMADA, AND Y. OTSUKA, Carbohydr. Res., 167 (1987) 221-237.
- 17 O. S. CHIZHOV, V. I. KADENTSEV, A. A. SOLOVYOV, P. F. LEVONOWICH, AND R. C. DOUGHERTY, J. Org. Chem., 41 (1976) 3425–3428.
- 18 N. K. KOCHETKOV AND O. S. CHIZHOV, Adv. Carbohydr. Chem., 21 (1966) 39-93.
- 19 S. SHIBUYA AND R. NAKANE, Phytochemistry, 23 (1984) 1425-1429.
- 20 M. A. JERMYN AND Y. M. YEOW, Aust. J. Plant Physiol., 2 (1975) 501-531.
- 21 A. G. DARVILL, M. McNeil, P. Albersheim, and D. P. Delmer, in N. E. Tolbert (Ed.), The Biochemistry of Plants, Academic Press, New York, 1980, pp. 91-162.
- 22 P. M. DEY AND K. BRINSON, Adv. Carbohydr. Chem. Biochem., 42 (1984) 265-382.
- 23 J. M. LAU, M. McNeil, A. G. DARVILL, AND P. ALBERSHEIM, Carbohydr. Res., 168 (1987) 245–274.
- 24 N. SHIMIZU AND M. TOMODA, Chem. Pharm. Bull., 31 (1983) 499-506.
- 25 H. YAMADA, T. NAGAI, J.-C. CYONG, Y. OTSUKA, M. TOMODA, N. SHIMIZU, AND K. SHIMADA, Carbohydr. Res., 144 (1985) 101–111.